

Study the antimicrobial effects of *Zataria multiflora*-based mouthwash on the microbial community of dental plaques isolated from children: A candidate of novel plant-based mouthwash

Estudio de los efectos antimicrobianos del enjuague bucal a base de Zataria multiflora sobre la comunidad microbiana de placas dentales aisladas de niños: Un candidato a enjuague bucal novedoso a base de plantas

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Abstract

Background: *Zataria multiflora* is a medicinal plant with high antimicrobial effects. It is mainly used as an oral specie in food. The present survey was aimed to assess the antimicrobial effects of *Z. multiflora*-based mouthwash on the microbial community of dental plaques.

Methods: Two-hundred dental plaque samples were collected from individuals. Culture technique was used to assess their microbial contamination. *Z. multiflora* was collected and use in a base of mouthwash in 1% concentration. Antimicrobial effects of *Z. multiflora*-based mouthwash was examined against isolated bacteria and compared to antibiotic agents using the disk diffusion. Minimum inhibitory concentration of *Z. multiflora*-based mouthwash was also studied against isolated bacteria.

Results: *Streptococcus mutans* (19%), *Enterobacter cloacea* (17.50%), and *Staphylococcus aureus* (15%) were the most commonly identified bacteria amongst the dental plaque samples. *H. pylori* (8%) had the lowest prevalence. The mean ranges of the diameter of the growth inhibition zones were 5.71 ± 0.92 (*S. aureus* against tetracycline) to 15.68 ± 0.55 (*S. mutans* against azithromycin) mm. *Z. multiflora* mouthwash (1%) harbored the highest antimicrobial effects against *S. mutans* (15.33 ± 0.81 mm), and *S. aureus* (12.01 ± 1.10 mm), while showed the lowest against *E. coli* (8.38 ± 0.46 mm) and *E. cloacea* (10.52 ± 0.84 mm). The lowest MIC levels were obtained for *S. mutans* (2 mg/ml). The highest MIC level was found for *E. cloacea* (8 mg/ml). The MIC levels of *Z. multiflora* mouthwash against *E. coli* and *H. pylori* bacteria were higher than examined concentrations.

Conclusion: *Z. multiflora*-based mouthwash may be a useful herbal-based mouthwash against bacteria in dental plaque samples.

Key words: *Zataria multiflora*, mouthwash, antimicrobial effects, dental plaques.

Resumen

Antecedentes: *Zataria multiflora* es una planta medicinal con altos efectos antimicrobianos. Se utiliza principalmente como especie oral en la alimentación. El presente estudio tenía como objetivo evaluar los efectos antimicrobianos de un enjuague bucal a base de *Z. multiflora* sobre la comunidad microbiana de las placas dentales.

Métodos: Se recogieron doscientas muestras de placa dental de individuos. Se utilizó una técnica de cultivo para evaluar su contaminación microbiana. Se recogió *Z. multiflora* y se utilizó en una base de enjuague bucal en una concentración del 1%. Se examinaron los efectos antimicrobianos del enjuague bucal a base de *Z. multiflora* contra las bacterias aisladas y se compararon con los agentes antibióticos mediante la difusión en disco. También se estudió la concentración mínima inhibitoria del enjuague bucal a base de *Z. multiflora* frente a bacterias aisladas.

Resultados: *Streptococcus mutans* (19%), *Enterobacter cloacea* (17,50%) y *Staphylococcus aureus* (15%) fueron las bacterias más comúnmente identificadas entre las muestras de placa dental. *H. pylori* (8%) tuvo la menor prevalencia. Los rangos medios del diámetro de las zonas de inhibición del crecimiento fueron de $5,71 \pm 0,92$ (*S. aureus* frente a la tetraciclina) a $15,68 \pm 0,55$ (*S. mutans* frente a la azitromicina) mm. El enjuague bucal de *Z. multiflora* (1%) albergó los mayores efectos antimicrobianos contra *S. mutans* ($15,33 \pm 0,81$ mm), y *S. aureus* ($12,01 \pm 1,10$ mm), mientras que mostró los más bajos contra *E. coli* ($8,38 \pm 0,46$ mm) y *E. cloacea* ($10,52 \pm 0,84$ mm). Los niveles de MIC más bajos se obtuvieron para *S. mutans* (2 mg/ml). El nivel de CIM más alto se encontró para *E. cloacea* (8 mg/ml). Los niveles de CIM de *Z. multiflora* contra las bacterias *E. coli* y *H. pylori* fueron superiores a las concentraciones examinadas.

Conclusión: El enjuague bucal a base de *Z. multiflora* puede ser un útil colutorio a base de plantas contra las bacterias presentes en las muestras de placa dental.

Palabras clave: *Zataria multiflora*, enjuague bucal, efectos antimicrobianos, placas dentales.

Introduction

Dental plaque is the name given to the aggregations of bacteria and their products which accumulate on the tooth surface¹. When plaque accumulates on the crowns of teeth the natural, smooth, shiny appearance of the enamel is lost and a dull, matt effect is produced². As it builds up, masses of plaque become more readily visible to the naked eye³. Additionally, dental plaque bacteria can be a dangerous sources of infections for other parts of the body, such as gastrointestinal tract, head, neck, and nasopharynx⁴.

In direct smears, the early plaque is dominated by cocci and rods, most of which are Gram-positive. In the mature plaque (after about 7 days) the percentage of cocci in the plaque decreases rapidly and filaments and rods constitute about 50% of organisms in plaque⁵. Studies revealed that *Streptococcus mutans*, *Enterobacter cloacea*, *Staphylococcus aureus*, *Klebsiella pneumonia*, *Helicobacter pylori*, and *Escherichia coli*, are the most important and frequent bacterial species isolated from the oral cavity and dental plaque samples, globally^{6,7}.

Several antimicrobial choices are available for the infections of the oral cavity. However, bacterial isolates of the dental plaque samples harbored severe resistance toward commonly used antimicrobial agents, particularly aminoglycosides, tetracyclines, penicillins, cephalosporins, and quinolones⁸. Thus, studying the profile and pattern of antibiotic resistance amongst bacterial isolates of dental plaques as novel reservoirs of bacteria seems essential.

Zataria multiflora (*Z. multiflora*) is a herbal plant belonging to the Lamiaceae family. It grows in Iran, Pakistan and Afghanistan and is known as Avishan Shirazi in Iran. The main components of the essential oil of this plant include phenolic compounds such as carvacrol, thymol and eugenol⁹. This plant is known as potential antimicrobial agents and mainly used as a spice in different foodstuffs¹⁰. Edible nature of this plant make it possible to use it in several types of oral drugs, particularly herbal mouthwash¹¹.

This study was aimed to assess the antimicrobial effects of *Z. multiflora*-based mouthwash on the microbial community of dental plaques as a novel candidate of plant-based mouthwash in vitro condition.

Materials and methods

Samples and inclusion criteria

A total of 200 children referred to the dentistry clinics for routine check-ups were assessed in this survey. All children with dental plaque samples were included in this survey. Dental plaque presence is the prominent inclusion factor. All children who had received antimicrobial options or antibacterial mouthwashes three months before the experiment were excluded from the research. Dental

plaque sample was taken from the gingival crevice at the most profound pocket reading and removed from the clinical site using a sterile universal curette. The curette tip was inserted into the depths of the crevice/pocket, moved coronally while in contact with the tooth surface to remove both sub and supragingival plaque.

Preparation of plant materials

Z. multiflora was purchased from traditional groceries. A total of 3500 grams of *Z. multiflora* was used to create an ethanol extract. It was cleaned, dried at room temperature for 24 hours, and processed in a blender until its texture became smooth. To obtain the extract from the processed *Z. multiflora*, 2000 ml of a 96% ethanol solution was used, and this process was repeated thrice. The initial extract was filtered and evaporated at temperatures ranging from 50 to 60 °C to obtain a 100% pure *Z. multiflora* extract. The pure extract was weighed, stored in a sealed glass container, and subsequently placed in a desiccator before being used as mouthwash.

Mouthwash formulation

As much as 100 ml of mouthwash was produced for each formulation with *Z. multiflora* extract as the active substance. The formulations of *Z. multiflora* mouthwash according to **table I**. Propylene glycol was included in the *Z. multiflora* extract and placed in a glass beaker. It was then raised to 60 °C, stirred with a magnetic stirrer at 300 rpm and Tween 80, and sorbitol and aquadest were added. Benzoic acid and sodium benzoate were dissolved in aquadest and added to the solution and stirred with a magnetic stirrer until homogeneous. Subsequently, 100 ml of the sorbitol and aquadest ad was stirred until the solution became clear, and Oleum menthae piperitae was added.

Table I: Formulation of *Z. multiflora* mouthwash.

Components	Frequency (%)
<i>Z. multiflora</i> ethanolic extract	1
Propylene glycol	25
Tween 80	5
Oleum menthae piperitae	0.25
Benzoate acid	0.1
Sodium benzoate	1
Sorbitol 70%	15
Aquadest	100

Quality assessment of *Z. multiflora* mouthwash

Organoleptic, acidity, stability, weight mass, viscosity, irritation, and contact time of *Z. multiflora* mouthwash were assessed using the method described previously¹².

Isolation of bacteria from the dental plaque samples

The dental plaque sample from each child was cultured into a sterile tube containing 5% sheep blood agar, chocolate agar and a selective medium and transported to the microbiology laboratory. All media were incubated at 37°C and 42 °C for 24 to 48 h. after Gram staining and

microscopy, different biochemical tests were performed to identify bacterial strains. The basic biochemical tests used to identify bacterial strains includes Starch Test, Simon Citrate, Oxidase, Catalase, Voges Proskauer, Urease, Indole, Methyl Red and Coagulase Test. Analytical Profile Index (API 20E) (BioMeriouxVitek, Inc., MO, USA) system was used to identify bacteria¹³.

Antibacterial effects of mouthwash against isolated bacteria

The simple disk diffusion method was used to assess the antimicrobial effects of synthesized mouthwash. For this purpose, isolated bacteria were cultured on Muller Hinton agar media. A total of 1000 µl of 1% *Z. multiflora* mouthwash were poured into the blank disk and located at the surface of each media. For comparison, penicillin (10 µg/disk), tetracycline (30 µg/disk), azithromycin (15 µg/disk), gentamicin (10 µg/disk), and ampicillin (10 µg/disk) (Oxoid, UK) antibiotic disks were accompanies. All guidelines were performed according to the Clinical and laboratory standard institute (CLSI)¹⁴⁻¹⁶. The Minimum Inhibitory Concentration (MIC) of synthetic *Z. multiflora* mouthwash was also assessed. For this purpose, 1, 2, 4, and 8 mg/ml concentration of mouthwash were prepared and the MIC value was determined using the previously described method¹⁷.

Statistical analysis

Collected data were transferred to the Microsoft Office Excel software and arranged well. Then, they were statistically analyzed using the SPSS software and chi-square and analysis of variance tests (ANOVA). $P < 0.05$ was consider z significant level¹⁸⁻²².

Results

Table II shows the distribution of bacteria strains isolated from dental plaque samples. As shown, *S. mutans* (19%), *E. cloacea* (17.50%), and *S. aureus* (15%) were the most commonly identified bacteria amongst the dental plaque samples. *H. pylori* (8%) had the lowest prevalence amongst the examined dental plaque samples. Statistically significant difference was obtained between the distribution of different bacteria ($P < 0.05$).

Table III shows the dimeter of the growth inhibition zone of bacteria against synthetic mouthwash compare to antimicrobial agents. The mean ranges of the diameter of the growth inhibition zones were 5.71 ± 0.92 (*S. aureus* against tetracycline) to 15.68 ± 0.55 (*S. mutans* against azithromycin) mm. *Z. multiflora* mouthwash (1%) harbored the highest antimicrobial effects against *S. mutans* (15.33 ± 0.81 mm), and *S. aureus* (12.01 ± 1.10 mm), while showed the lowest against *E. coli* (8.38 ± 0.46 mm) and *E. cloacea* (10.52 ± 0.84 mm). Statistically significant differences were obtained between the diameter of the growth inhibition zone of bacteria treated with different antimicrobial agents ($P < 0.05$).

Table IV shows the MIC values of *Z. multiflora* mouthwash against different isolated bacteria. Findings showed that the lowest MIC levels were obtained for *S. mutans* (2 mg/ml). The highest MIC level was found for *E. cloacea* (8 mg/ml). The MIC levels of *Z. multiflora* mouthwash against *E. coli* and *H. pylori* bacteria were higher than examined concentrations (non detected).

Table II: Distribution of bacteria strains isolated from dental plaque samples.

Samples	N. collected	Distribution of bacteria (%)				
		<i>S. aureus</i>	<i>S. mutans</i>	<i>E. cloacea</i>	<i>E. coli</i>	<i>H. pylori</i>
Dental plaques	200	30 (15)	38 (19)	35 (17.50)	20 (10)	16 (8)

Table III: Dimeter of the growth inhibition zone of bacteria against synthetic mouthwash compare to antimicrobial agents.

Tested antimicrobial agents	Diameter of the growth inhibition zone of bacteria (mm)				
	<i>S. aureus</i>	<i>S. mutans</i>	<i>E. cloacea</i>	<i>E. coli</i>	<i>H. pylori</i>
<i>Z. multiflora</i> mouthwash (1%)	12.01 ± 1.10^a	15.33 ± 0.81^a	10.52 ± 0.84^a	8.38 ± 0.46^b	10.57 ± 0.61^a
Penicillin	8.63 ± 0.34^b	9.66 ± 0.23^c	10.14 ± 0.22^a	8.15 ± 0.51^b	10.33 ± 0.35^a
Tetracycline	5.71 ± 0.92^c	6.61 ± 0.28^d	8.17 ± 0.37^b	8.81 ± 0.36^b	8.66 ± 0.25^a
Azithromycin	13.24 ± 0.95^a	15.68 ± 0.55^a	11.43 ± 0.52^a	10.93 ± 0.44^a	9.90 ± 0.60^a
Gentamicin	6.03 ± 0.32^c	11.82 ± 0.39^b	11.97 ± 0.93^a	8.55 ± 0.81^b	9.93 ± 0.65^a
Ampicillin	7.15 ± 0.41^c	10.17 ± 0.16^c	10.08 ± 0.09^a	9.72 ± 0.74^{ab}	9.71 ± 0.69^a

*Dissimilar small letters in each column show significant statistical differences ($P < 0.05$).

Table IV: MIC values of *Z. multiflora* mouthwash against different isolated bacteria.

Treatment	MIC (mg/ml)				
	<i>S. aureus</i>	<i>S. mutans</i>	<i>E. cloacea</i>	<i>E. coli</i>	<i>H. pylori</i>
<i>Z. multiflora</i> mouthwash	4	2	8	ND*	ND

*Non detected.

Discussion

Infections may cause several life-threatening diseases globally²³⁻³⁸. In this regard, medical plants and traditional medicine act as healing sciences³⁹. In this study, *Z. multiflora*-based mouthwash was used as an antimicrobial agent on bacteria isolated from dental plaque samples. Total distribution of *S. aureus*, *S. mutans*, *E. cloacea*, *E. coli*, and *H. pylori* amongst the examined dental plaque samples was 15%, 19%, 17.50%, 10%, and 8%, respectively.

An Indian survey⁴⁰ revealed that *Streptococcus* spp. was the most commonly detected bacteria (51.00%), followed by *E. coli* (19.00%) and *Veillonella* spp. (19.00%). A research on United Kingdom⁴¹ showed that *Tannerella forsythensis* (65.00%), *Porphyromonas gingivalis* (49.00%), and *Actinobacillus actinomycetemcomitans* (55.00%) were the most commonly detected pathogens isolated from dental plaque samples. Similarly in Korea⁴², *Streptococcus*, *Corynebacterium*, *Neisseria*, and *Fusobacterium* were the most commonly detected bacterial strains in dental plaque samples. Brazilian survey⁴³ showed that the predominant species in all 600 samples included *Corynebacterium diphtheriae*, *Enterococcus faecalis*, *S. aureus*, *Acinetobacter baumannii*, *P. aeruginosa*, and *E. coli* were predominant bacterial pathogens in the dental plaque samples of children with chronic periodontitis.

Findings of the current research revealed that *Z. multiflora*-based mouthwash had the highest antimicrobial effects on *S. mutans* and *S. aureus*. The main reason for the lower antimicrobial effects of the mouthwash against other bacteria is that they were Gram-negative and have Lipopolysaccharide (LPS) in their cell walls which inhibit the penetration of essential oils and herbal extracts. Aghili et al. (2015)⁴⁴ reported that the *Z. multiflora* extract had the higher antimicrobial effects on experimentally contaminated orthodontic elastomeric ligatures compared to chlorhexidine mouthwash. Milho et al. (2021)⁴⁵ stated that the essential oils of *Cymbopogon citratus* (DC.) Stapf and *Lippia alba* (Mill.) seem to be the most promising in fighting microbial biofilm in *S. mutans*, given their high capacity to reduce biofilm at low concentrations. Significant effects of herbal mouthwash against pathogenic bacteria recovered from the dental plaques have similarly been reported by Tusi et al. (2020) (Iran)⁴⁶ and Pedrazzi et al. (2015) (Brazil)⁴⁷.

Conclusion

According to the high antimicrobial effects of the *Z. multiflora*-based mouthwash compared to antimicrobial agents even in low concentrations, its application as a novel and herbal-based mouthwash particularly against Gram-positive bacteria responsible for dental plaque formation has been recommended. However, some additional surveys should perform to assess other effects of *Z. multiflora*-based mouthwash on the oral cavity.

Conflict of Interest

The authors declare that they have no conflict of interest.

References

- Murakami S, Mealey BL, Mariotti A, Chapple IL. Dental plaque-induced gingival conditions. *Journal of clinical periodontology*. 2018 Jun;45:S17-27.
- Sreenivasan PK, Prasad KV, Javali SB. Oral health practices and prevalence of dental plaque and gingivitis among Indian adults. *Clinical and experimental dental research*. 2016 Jun;2(1):6-17.
- Marsh PD. Dental plaque as a biofilm and a microbial community—implications for health and disease. In *BMC Oral health 2006 Jun (Vol. 6, No. 1, pp. 1-7)*. BioMed Central.
- Li X, Koltveit KM, Tronstad L, Olsen I. Systemic diseases caused by oral infection. *Clinical microbiology reviews*. 2000 Oct 1;13(4):547-58.
- Selwitz RH, Ismail AI, Pitts NB. Dental caries. *The Lancet*. 2007 Jan 6;369(9555):51-9.
- Marsh PD. Dental plaque as a biofilm and a microbial community—implications for health and disease. In *BMC Oral health 2006 Jun (Vol. 6, No. 1, pp. 1-7)*. BioMed Central.
- Marsh PD. Dental plaque as a microbial biofilm. *Caries research*. 2004;38(3):204-11.
- Meinen A, Reuss A, Willrich N, Feig M, Noll I, Eckmanns T, Al-Nawas B, Markwart R. Antimicrobial Resistance and the Spectrum of Pathogens in Dental and Oral-Maxillofacial Infections in Hospitals and Dental Practices in Germany. *Frontiers in microbiology*. 2021 Jun 2;12:1418.
- Dini S, Dadkhah A, Fatemi F. Biological Properties of Iranian Zataria Multiflora Essential Oils: A Comparative Approach. *Electronic Journal of Biology*. 2015;11(3):57-62.
- Aghili H, Nadoushan AA, Herandi V. Antimicrobial effect of zataria multiflora extract in comparison with chlorhexidine mouthwash on experimentally contaminated orthodontic elastomeric ligatures. *Journal of dentistry (Tehran, Iran)*. 2015 Jan;12(1):1.
- Owlia P, Pirveicy H, Saderi H, Rezvani MB, Mansouri S. Evaluation of the antimicrobial effects of extract of Zataria multiflora against oral Streptococci. *Iranian Journal of Pharmaceutical Research*. 2010 Nov 20(Supplement 2):74-5.
- Iskandar B, Lukman A, Syaputra S, Al-Abrori UN, Surboyo MD, Lee CK. Formulation, characteristics and anti-bacterial effects of Euphorbia hirta L. mouthwash. *Journal of Taibah University Medical Sciences*. 2021 Sep 17.
- Shayegani ME, Maupin PS, McGlynn DM. Evaluation of the API 20E system for identification of nonfermentative Gram-negative bacteria. *Journal of Clinical Microbiology*. 1978;7(6):539-45.
- Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing; twenty-second informational supplement. M100-S21. Wayne Pa: CLSI; 2012.
- Ranjbar R, Seif A, Dehkordi FS. Prevalence of antibiotic resistance and distribution of virulence factors in the shiga toxigenic *Escherichia coli* recovered from hospital food. *Jundishapur Journal of Microbiology*. 2019;12(5):8.
- Rahi A, Kazemeini H, Jafariaskari S, Seif A, Hosseini S, Dehkordi FS. Genotypic and phenotypic-based assessment of antibiotic resistance and profile of staphylococcal cassette chromosome mec in the methicillin-resistant *Staphylococcus aureus* recovered from raw milk. *Infection and drug resistance*. 2020;13:273.
- Dadashi M, Hashemi A, Eslami G, Fallah F, Goudarzi H, Erfanimanesh S, et al. Evaluation of antibacterial effects of Zataria multiflora Boiss extracts against ESBL-producing *Klebsiella pneumoniae* strains. *Avicenna journal of phytomedicine*. 2016 May;6(3):336.
- Ranjbar R, Yadollahi Farsani F, Safarpour Dehkordi F. Antimicrobial resistance and genotyping of *vacA*, *cagA*, and *iceA* alleles of the *Helicobacter pylori* strains isolated from traditional dairy products. *Journal of Food Safety*. 2019 Apr;39(2):e12594.
- Dehkordi FS, Momtaz H, Doosti A. Application of Real-Time PCR for detection of *Aspergillus* species in aborted ruminant fetuses. *Bulg. J. Vet. Med*. 2012 Mar 1;15(1):30-6.
- Dehkordi FS. Prevalence study of *Coxiella burnetii* in aborted ovine and caprine fetuses by evaluation of nested and real-time PCR assays. *American Journal of Animal and Veterinary Sciences*. 2011.
- Nejat S, Momtaz H, Yadegari M, Nejat S, Safarpour Dehkordi F, Khamesipour F. Seasonal, geographical, age and breed distributions of equine viral arteritis in Iran. *Kafkas Univ Vet Fak Derg*. 2015 Jan 1;21(1):111-6.
- Dehkordi FS, Saberian S, Momtaz H. Detection and segregation of *Brucella abortus* and *Brucella melitensis* in Aborted Bovine, Ovine, Caprine, Buffaloes and Camelid Fetuses by application of conventional and real-time polymerase chain reaction. *The Thai Journal of Veterinary Medicine*. 2012 Mar 1;42(1):13.
- Sheikhshahrokh A, Ranjbar R, Saeidi E, Dehkordi FS, Heiat M, Ghasemi-Dehkordi P, et al. Frontier therapeutics and vaccine strategies for sars-cov-2 (COVID-19): A review. *Iranian Journal of Public Health*. 2020 Jul 11.
- Dehkordi FS, Khamesipour F, Momeni M. *Brucella abortus* and *Brucella melitensis* in Iranian bovine and buffalo semen samples: The first clinical trial on seasonal, Senile and geographical distribution using culture, conventional and real-time polymerase chain reaction assays. *Kafkas Univ Vet Fak Dergisi*. 2014;20(6):821-.
- Dehkordi FS. Prevalence study of Bovine viral diarrhea virus by evaluation of antigen capture ELISA and RT-PCR assay in Bovine, Ovine, Caprine, Buffalo and Camel aborted fetuses in Iran. *AMB express*. 2011 Dec;1(1):1-6.
- Dehkordi FS, Haghighi N, Momtaz H, Rafsanjani MS, Momeni M. Conventional vs real-time PCR for detection of bovine herpes virus type 1 in aborted bovine, buffalo and camel fetuses. *Bulgarian Journal of Veterinary Medicine*. 2013 Jun 1;16(2).
- Dehkordi FS, Valizadeh Y, Birgani TA, Dehkordi KG. Prevalence study of *Brucella melitensis* and *Brucella abortus* in cow's milk using dot enzyme linked immuno sorbent assay and duplex polymerase chain reaction. *J Pure Appl Microbiol*. 2014;8(2):1065-9.
- Abdolmaleki Z, Mashak Z, Safarpour Dehkordi F. Molecular and virulence characteristics of methicillin-resistant *Staphylococcus aureus* bacteria recovered from hospital cockroaches. *Jundishapur Journal of Microbiology*. 2019 Dec 31;12(12).
- Mashak Z, Jafariaskari S, Alavi I, Shahreza MS, Dehkordi FS. Phenotypic and genotypic assessment of antibiotic resistance and genotyping of *vacA*, *cagA*, *iceA*, *oipA*, *cagE*, and *babA2* alleles of *Helicobacter pylori* bacteria isolated from raw meat. *Infection and drug resistance*. 2020;13:257.
- Dehkordi FS, Haghighi Borujeni MR, Rahimi E, Abdizadeh R. Detection of *Toxoplasma gondii* in raw caprine, ovine, buffalo, bovine, and camel milk using cell cultivation, cat bioassay, capture ELISA, and PCR methods in Iran. *Foodborne Pathogens and Disease*. 2013 Feb 1;10(2):120-5.

31. Dehkordi FS, Gandomi H, Basti AA, Misaghi A, Rahimi E. Phenotypic and genotypic characterization of antibiotic resistance of methicillin-resistant *Staphylococcus aureus* isolated from hospital food. *Antimicrobial Resistance & Infection Control*. 2017 Dec;6(1):1-1.
32. Dehkordi FS, Yazdani F, Mozafari J, Valizadeh Y. Virulence factors, serogroups and antimicrobial resistance properties of *Escherichia coli* strains in fermented dairy products. *BMC research notes*. 2014 Dec;7(1):1-8.
33. Dehkordi FS, Parsaei P, Saberian S, Moshkelani S, Hajshafiei P, Hoseini SR, et al. prevalence study of *Theileria annulata* by comparison of four diagnostic techniques in southwest Iran. *Bulgarian Journal of Veterinary Medicine*. 2012 Jun 1;15(2).
34. Dehkordi FS, Barati S, Momtaz H, Ahari SN, Dehkordi SN. Comparison of shedding, and antibiotic resistance properties of *Listeria monocytogenes* isolated from milk, feces, urine, and vaginal secretion of bovine, ovine, caprine, buffalo, and camel species in Iran. *Jundishapur Journal of Microbiology*. 2013 May 1;6(3):284.
35. Dehkordi FS, Valizadeh Y, Birgani TA, Dehkordi KG. Prevalence study of *Brucella melitensis* and *Brucella abortus* in cow's milk using dot enzyme linked immuno sorbent assay and duplex polymerase chain reaction. *J Pure Appl Microbiol*. 2014;8(2):1065-9.
36. Dehkordi FS, Rafsanjani MS. Prevalence study of *Coxiella burnetii* in aborted fetuses of small ruminants in various partum and seasons in Iran. *African Journal of Microbiology Research*. 2012 Jul 19;6(27):5594-600.
37. Dehkordi FS, Tavakoli-Far B, Jafariaskari S, Momtaz H, Esmailzadeh S, Ranjbar R, et al. Uropathogenic *Escherichia coli* in the high vaginal swab samples of fertile and infertile women: virulence factors, O-serogroups, and phenotyping and genotyping characterization of antibiotic resistance. *New Microbes and New Infections*. 2020 Nov 1;38:100824.
38. Safarpordehkordi F, Yahaghi E, Khodaverdi Darian E. Prevalence of antibiotic resistance in *Escherichia coli* isolated from poultry meat supply in Isfahan. *Iranian Journal of Medical Microbiology*. 2014 Aug 10;8(2):41-7.
39. Dehkordi FS, Tirgir F, Valizadeh Y. Effects of Guajol® ointment synthesized from medicinal smoke condensate of jennet feces on burn wound healing on Wistar rat. In *Veterinary Research Forum 2017* (Vol. 8, No. 3, p. 215). Faculty of Veterinary Medicine, Urmia University, Urmia, Iran.
40. Srivastava A, Saha S, Sahu C. Early and accurate detection of bacterial isolates from dental plaque in subjects with primary, mixed, and permanent dentition by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry technique. *Journal of Indian Society of Periodontology*. 2020 Mar;24(2):104.
41. Gafan GP, Lucas VS, Roberts GJ, Petrie A, Wilson M, Spratt DA. Prevalence of periodontal pathogens in dental plaque of children. *Journal of Clinical Microbiology*. 2004 Sep;42(9):4141-6.
42. Lee E, Park S, Um S, Kim S, Lee J, Jang J, et al. Microbiome of Saliva and Plaque in Children According to Age and Dental Caries Experience. *Diagnostics*. 2021 Aug;11(8):1324.
43. Souto R, Andrade AF, Uzeda M, Colombo AP. Prevalence of "non-oral" pathogenic bacteria in subgingival biofilm of subjects with chronic periodontitis. *Brazilian Journal of Microbiology*. 2006;37:208-15.
44. Aghili H, Nadoushan AA, Herandi V. Antimicrobial effect of *zataria multiflora* extract in comparison with chlorhexidine mouthwash on experimentally contaminated orthodontic elastomeric ligatures. *Journal of dentistry (Tehran, Iran)*. 2015 Jan;12(1):1.
45. Milho C, Silva J, Guimarães R, Ferreira IC, Barros L, Alves MJ. Antimicrobials from Medicinal Plants: An Emergent Strategy to Control Oral Biofilms. *Applied Sciences*. 2021 Jan;11(9):4020.
46. Tusi SK, Jafari A, Marashi SM, Niknam SF, Farid M, Ansari M. The effect of antimicrobial activity of *Teucrium Polium* on Oral *Streptococcus Mutans*: a randomized cross-over clinical trial study. *BMC Oral Health*. 2020 Dec;20(1):1-8.
47. Pedrazzi V, Leite MF, Tavares RC, Sato S, do Nascimento GC, Issa JP. Herbal mouthwash containing extracts of *Baccharis dracunculifolia* as agent for the control of biofilm: clinical evaluation in humans. *The Scientific World Journal*. 2015 Jan 1;2015.